

# New Genetic and Linguistic Analyses Show Ancient Human Influence on Baobab Evolution and Distribution in Australia

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## Abstract

This study investigates the role of human agency in the gene flow and geographical distribution of the Australian baobab *Adansonia gregorii*. The genus *Adansonia* is a charismatic tree endemic to Africa, Madagascar, and northwest Australia. It has long been valued by humans for its multiple uses. The distribution of genetic variation in baobabs in Africa has been attributed to human-mediated dispersal over millennia, but this relationship has never been investigated for the Australian baobab. We combined genetic and linguistic data to analyse geographic patterns of gene flow and movement of word-forms in the Aboriginal languages of northwest Australia. Comprehensive assessment of genetic diversity showed weak geographic structure and high gene flow. Of potential dispersal vectors, humans were identified as most likely to have enabled dispersal across biogeographic barriers in northwest Australia. Genetic-linguistic analysis demonstrated congruence of gene and directional movement of Aboriginal loanwords for *A. gregorii*. These findings, along with previous archaeobotanical data from the Late Pleistocene and Holocene, suggest that ancient humans significantly influenced the geographic distribution of *Adansonia* in northwest Australia.

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## Introduction

The role of humans in shaping crop genetic diversity has always been considered an integral factor in the evolution of various regions of the world [1–3]. Interdisciplinary research combining genetics, linguistics, and archaeobotany has enhanced understanding of the geographic patterns of animal and crop domestication and subsequent diffusion by humans. Yet, there is very little comparable research on how anthropogenic agency has influenced the evolution and distribution of uncultivated plants that, nonetheless, have a long history of human use [4–6]. A striking example is that of the Australian baobab tree, an iconic genus endemic to Africa, Madagascar, and the Kimberley region of northwest Australia [7,8]. These trees hold significant cultural symbolism and multipurpose value as sources of food, medicine, water storage, shelter, and material for artisanal products in all these places [9–17]. Although there is no evidence of baobabs being cultivated in Australia, the distribution of the African baobab species, *Adansonia digitata* L., has been closely linked to human dispersal and settlement patterns [18,19]. This association is also recognised by the diversity and borrowing of terms for baobabs between languages in Africa [4, 16]. In contrast, previous research on the evolution and geographic distribution of the Australian baobab *gregorii* F. Muell., has been based on the assumption of long-term natural processes [7,8] without any significant influence of human agency. This assumption may have stemmed from the long-held view of Aboriginal Australia as a 'continent

gatherers' [20–24] where anthropogenic agency was limited to 'fire-stick farming' of landscapes for nomadic foraging [25,26]. We explore the role of humans in shaping the evolution of *A. gregorii* by determining whether the geographic genetic diversity is explained, in part, by patterns of human migration, as inferred from linguistic analysis.

Levels of genetic divergence show that *A. gregorii* separated from other *Adansonia* species more recently than the Gondwana, but before the arrival of humans in Australia [7,8]. Leong-Pock Tsy et al. [27] demonstrated that *A. digitata* their viability in seawater, making oceanic current dispersal feasible. From this it can be inferred that *A. gregorii* has been in Australia for longer than humans. There is also the possibility, albeit a less parsimonious explanation, that the species arrived recently from an unknown population which is now extinct. One hypothesis outlining how *A. gregorii* may have arrived with humans has been explored in more detail by Pettigrew [28].

*Adansonia gregorii*, known in Australia as 'boab', is mainly distributed across the Kimberley region of northwest Australia, with a small extension eastward into the Victoria River District in the Northern Territory. The distribution of *A. gregorii* in Australia extends from the northern coastline to the edge of the Great Sandy Desert and the Tanami Desert [17,29–32]. The region represents the westernmost extent of the Australian Monsoon Tropics (AMT), which is characterised by high rainfall and savanna vegetation [17,33]. The tree has been introduced more recently in urban centers of northern Australia for ornamental purposes [17].

The AMT biome is bounded to the south by arid habitats, which began developing in the Late Cenozoic and contain different biota [34,35]. The major biogeographical divide in northwest Australia is between the Kimberley to the west and the Land to the east, with more localized and specific barriers created by major river drainage systems [36–39]. Within phylogeographic patterns for rock-wallabies (*Petrogale* spp.) and other species suggest an East-West Divide running through Central Kimberley [34,37,39]. Despite evidence of biogeographic barriers, a previously detailed population genetic analysis of *A. gregorii* has demonstrated that there is little genetic structure, with  $F_{ST}$  values non-significant between most populations. A geographic structure could be explained by a relatively recent arrival in the Kimberley, a recent genetic bottleneck, or high dispersal rates across the species' range. For reasons detailed in Bell et al. [40], high dispersal is the most likely explanation.

In this paper we sought to evaluate the latter hypothesis – that the low levels of genetic structure within *A. gregorii* are due to levels of gene flow and, specifically, that human-mediated seed dispersal has been an important evolutionary factor for this species. Pollination in *A. gregorii* might occasionally entail birds or bats, but it appears that hawkmoths are the primary agents [17,29,33,41,42]. Pollination-mediated gene flow is limited to the paternal genome and, in insect pollinated species, a less effective mechanism of long-distance gene flow than fruit dispersal [43–45]. Floodwaters could explain some of the observed patterns [7,16], but would probably not spread seeds beyond the edges of seasonal waterways and alluvial flats due to the indehiscent nature of the *A. gregorii* pericarp [7,33]. Other seed dispersing agents could be mammals such as rock wallabies (*Petrogale* spp.), other wallabies and kangaroos (*Macropus* spp.), which eat the fruit and disperse the seeds in their droppings. However, phylogeographic studies of the short-eared rock-wallaby (*P. brachyotis*), showed strong genetic structure, suggesting that at least this species has limited dispersal ability across biogeographic barriers in the Kimberley [39,46].

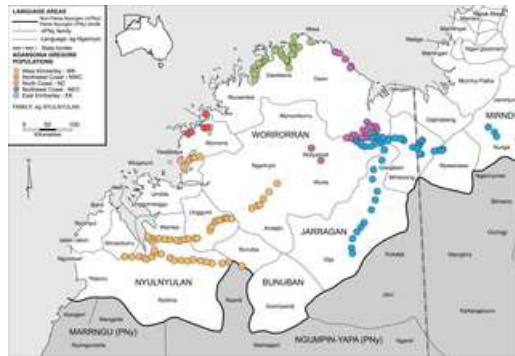
Anthropogenic agency has not been formally considered as a gene flow vector for *A. gregorii*, despite archaeological evidence of long-term use by Aboriginal groups in the region [28,47–50]. This omission may be due to the fact that the boab was not historically, or not considered part of the food crops traditionally harvested by Aboriginal groups. Also, the presence of a boab pod remains at one or two archaeological sites is not sufficient to demonstrate anthropogenic dispersal across the species' geographic range. Additional evidence is needed to evaluate whether humans have played a role in boab gene flow. Historical linguistics, which has been used in combination with other sources of data to trace geographic patterns in domesticated species [1–3], could be applied to a non-domesticated tree species such as *A. gregorii* [6]. Specifically, we investigate the role of human agency in the gene flow of *A. gregorii* by testing for congruence between the spatial patterns of genetic variation in *A. gregorii* trees and associated word-forms in the Aboriginal languages of northwest Australia. A high level of spatial overlap between the genetic and linguistic data. Our results indicate that, as previously shown for *A. digitata* in Africa [19,51,52], ancient humans have influenced the distribution of genetic diversity of *A. gregorii* in northern Australia by acting as seed dispersal agents over long distances.

#### Materials and Methods

This study makes use of two recently published data sets relating to *A. gregorii* from different fields of research. The first is genetic data from six nuclear microsatellite loci, of 220 *A. gregorii* individuals [40] (S1 Dataset). Although the number of microsatellite loci is low and can render some quantitative methods inaccurate, we used this data to make qualitative inferences on patterns of dispersal and relationship between populations. The second data set consists of the words used for 'boab' in 11 Aboriginal languages from across the species range [53] (S2 Dataset). This study brings these two datasets together and analyses them simultaneously to detect any congruent patterns.

#### Phylogenetic analysis of microsatellite data

We conducted phylogenetic analysis of the *A. gregorii* populations shown in Fig. 1, using previously obtained microsatellite data [40]. Collection of plant material, with appropriate permissions, deposition of specimens, and laboratory analyses as detailed in Bell et al. [40]. This study used the spatial principal components analysis (sPCA) to map and define genetic populations. We identified five genetically differentiated clusters: West Kimberley (WK); East Kimberley (EK); North West Coast (NWC); North Central (NC), and North East Coast (NEC). Genetic divergence between these clusters was weak, but statistically significant. Therefore, these were treated as five populations for subsequent analyses.



**Fig 1. Graphical representation of the five inferred *Adansonia gregorii* populations from the sPCA, shown on a map of Aboriginal areas of the Kimberley region.**

Map based on Harvey [58] and McGregor and Rumsey [63].

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To obtain an estimate of the relationships among the five populations, we first obtained 1000 bootstrap replicates of microsatellite data. We calculated pairwise genetic distances between populations with the  $D_C$  method [54] for each replicate in Phylip v 3.69 [55]. These distance matrices were then used to construct unrooted neighbour-joining tree bootstrap consensus tree in Phylip. While more complex methods have been found to provide greater accuracy in divergence events from microsatellite data, the simple  $D_C$  method was judged suitable because it performs as well as other methods at phylogenetic reconstruction [56]. The resulting tree was midpoint rooted.

#### Phylogeny of linguistic data

The Aboriginal languages of Australia are broadly classified into a number of families, Pama-Nyungan (**PNy**) and non-Pama-Nyungan (**nPNy**) families [57,58]. The **PNy** language groups are most widespread across the continent, with Pama-Nyungan families (excluding Tasmania) exist primarily in northwest Australia across the Kimberley and western Arnhem Land. **PNy** language subgroups and **nPNy** families have been distinguished by the comparative method [60], including grammatical morphology and measures of difference in vocabulary [57], with sub-groups identified by sets of 'shared innovations'. Neighbour Joining analysis of twenty-one Kimberley languages using the method described in Hudson and Bryant [59] published by McGregor and Rumsey [63]. The tree generated from that analysis showed the relationships between languages based on lexical resemblances from a basic wordlist of 105 meanings, containing minimum numbers of 10 words. A greater degree of proximity and shared branches in the phylogeny indicates higher lexical similarity between the languages. We used this neighbour-joining tree in conjunction with a more general consensus about family classification comparative method in discussion of the origins of words for the boab tree [64].

#### Determining boab proto-words, inherited forms, and loanwords

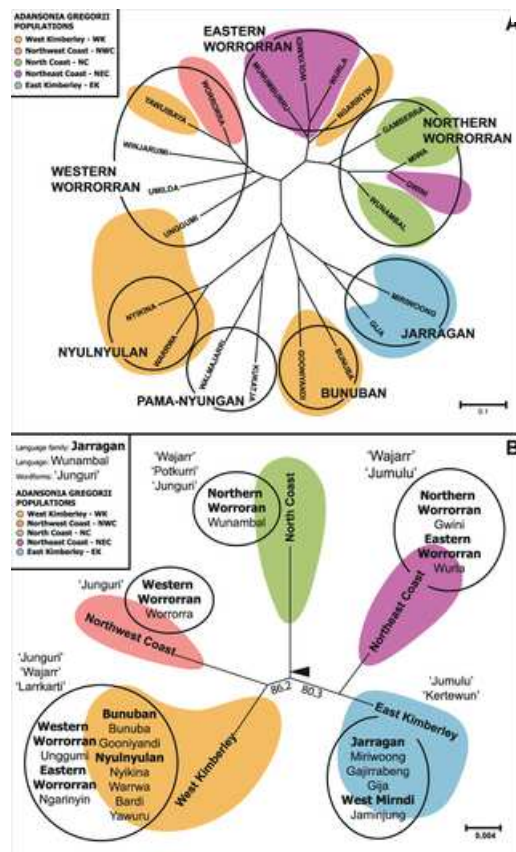
Data on boab word-forms and related terminology was drawn from both published and unpublished sources and translated to a standard orthography [53] (S2 Dataset). The distinction between inheritance (vertical transmission) and borrowing (horizontal transmission) of word-forms is vital to our analysis. Proto-words are word-forms that can be traced back through plausible steps to a common ancestral word-form in an ancestral language (known as the proto-language) [60]. In vertical transmission, proto-words are inherited through nodes in a phylogeny to a number of more recent descendant languages, changing meaning and form over time. Horizontal transmission, or diffusion, occurs through borrowing, whereby words become adopted in other languages that may or may not be closely related by inheritance. There are several types of methods used to detect loanwords and their original source [6]. Borrowed words in a language are ones that cannot be connected to plausible proto-words in that language but instead show strong similarity to words in another language. The identification of a loanword is further corroborated when the word can be analyzed into meaningful word-parts in the other language (the loan source) but not in a language into which it has been borrowed.

#### Genetic-Linguistic Analysis

The spatial distribution of each language was defined following Harvey [58] and McGregor and Rumsey [63]. The *A. gregorii* distribution includes five **nPNy** families (Worrorran, Nyulnyulan, Bunuban, Jarragan, and Mirndi) and two **PNy** languages (Marrngu and Ngumpin-Yapa) (Fig. 1). Three sets of relationships of genetic and linguistic data for *A. gregorii* were used as evidence of human-mediated dispersal across the plant's geographic range.

#### Boab populations and language families.

Geographic congruence between *A. gregorii* populations and language areas was examined by superimposing the spatial distribution of the phylogenetic tree of the main Kimberley language groups analysed by McGregor and Rumsey [63], and conversely superimposing the language group areas [58] in which the boab populations occur on the neighbour-joining tree of *A. gregorii* populations (Fig. 2A and B). A high level of geographic congruence between the *A. gregorii* genetic populations and the language areas they occupy would be consistent with the idea that people have moved boabs extensively within language areas and much between language areas.



**Fig 2. A. Neighbour-joining tree of lexical resemblance among Worrوران and nearby Aboriginal languages of the Kimberley, foll McGregor and Rumsey [63].**

Twenty-one languages (19 belonging to *nPNy* families and 2 belonging to the Ngumpin *PNy* sub-group) were ch analysis on the basis of availability of relatively reliable information. Distance between nodes indicates degree of between the languages according to the scale shown. The five inferred *A. gregorii* populations have been superi the tree to visualize spatial congruence with language groups. **B. Neighbour-joining phylogeny of the five inf *gregorii* populations showing their occurrence in language group areas according to Harvey [58].** Distanc nodes indicates genetic divergence between populations calculated with the  $D_C$  method. Arrow indicates mid-po word-forms for boabs associated with each of the five inferred *A. gregorii* populations are shown in quotation ma doi:10.1371/journal.pone.0119758.g002

Analysis of molecular variance (AMOVA [65]) was used to test whether there was a significant broad scale correlati genetics and language family area. Boab individuals were grouped according to their occurrence in the nPNy family significance was tested with 999 permutations.

A partial Mantel test of genetic distance vs. language family area with geographic distance as a covariate [66] was whether any statistical significance inferred by the AMOVA was a result of isolation by distance in both language ar variation. Genetic distance was calculated between boab individuals using the  $D_C$  method [54], and boab individual to language family areas as described above.

We used a permutation test, implemented in Mesquite [67], to test for statistically significant association between la and genetic identity. The genetic clusters were scored as single, multistate characters. The length of this character phylogenetic tree was calculated under equally-weighted parsimony. We permuted this character 999 times, and ca length on the language phylogenetic tree. We then determined whether the length of the original character was insi distribution based on the randomly permuted character.

Language families and boab word-forms.

To examine the geographic relationship between boab word-forms and language families, the numbers of boab wor of the *nPNy* language families were examined to identify languages with higher and lower numbers of boab word-fr greater diversity of word-forms for *A. gregorii* within language family areas may suggest long-term presence of, and boab populations. Conversely, lower diversity of word-forms for the tree could be either due to recent rapid expansi family area, or to a more recent interaction of those populations with boabs.

[illegible]

**Table 1. Word-forms for *Adansonia gregorii* in the Kimberley region.**  
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Data on boab word-forms and related terminology were drawn from both published and unpublished sources (S2 D left-hand columns identify the non-Pama-Nyungan language family and Pama-Nyungan (*PNy*) sub-groups, and the language groups. The word-forms are represented in a standardized orthography. A hyphen indicates a morpheme separating a stem from a suffix. The subsequent columns list the reconstructed or loan-source form and the probable meaning of the reconstructed source form. Section A lists word-forms for the boab tree; Section B lists word-forms for tree-parts.

Boab word-forms and genetic population phylogeny.

If humans have played a major role in dispersing boabs over the Kimberley, then we might expect congruence between loanwords and dispersal of genotypes. Gene flow between *A. gregorii* populations has previously been examined by coalescent analysis, using IMA2 [40]. That analysis yielded estimates of the relative migration rates in two major directions between western and eastern populations and between coastal and inland populations [40]. These directional patterns were compared with the directional movement of boab word-forms, especially loanwords, using the comparative method to trace their original source and examine patterns of geographic congruence. The reconstructed loan source forms for each word in Table 1 were used to identify five words which showed directional movement of borrowing between the language and the Kimberley region.

## Results

### Phylogeny of *A. gregorii* populations

Phylogenetic analysis generated an unrooted tree for the five inferred populations with bootstrap support values greater than 50% for both internal branches (Fig. 2B). Midpoint rooting suggests that the WK and NWC populations are sisters, as are the NC and NEC populations, with the deepest divergence separating the NC population from the other four. However, since the node is not supported by bootstrap values, this conclusion should be treated with caution.

### Boab populations and language family areas

The association of the five *A. gregorii* populations on the language phylogenetic tree (Fig. 2A) and vice-versa, of lar with boab population phylogeny (Fig. 2B), did not reveal strict geographic congruence. Statistical analyses showed variance between boab individuals from different *nPNy* family areas was low but statistically significant (AMOVA, 3<sup>rd</sup> genetic variation,  $P = 0.001$ ). A partial Mantel test of genetic distance vs. language family area with geographic distance as covariate was not significant ( $R_{xy} = 0.00728$ ;  $P = 0.285$ ), indicating that the statistical significance of the AMOVA co spatial autocorrelation in both languages and genetic variation. However, it is noteworthy that there seems to be a sharp distinction between the EK and NEC genetic clusters, and this break coincides with the distinction between the Worrorrnan language families.

The mapping of genetic population assignments onto the language tree required 4 steps for the original, unpermuted the permuted characters required from 5 to 10 steps, with a mean of 8.55. With 999 permutations, this implies a p-value indicating a strongly non-random pattern of association between the language tree and the genetic clusters. Because geography was not included as a covariate, we cannot rule out the possibility that this association is driven by geography rather than the relationship between boab genetics and human language variation.

#### Language family areas and *A. gregorii* word-forms

The greatest diversity of word-forms for *A. gregorii* is found in the northern coastal areas encompassing the Worror family, followed by those in the Jarragan family (Table 1 and Fig. 2A). Each has one dominant word for the tree species borrowed from elsewhere. Worrorran has the term *junguri*, and Jarragan has *jumulu*. Both terms are reconstructable for boab in these language families. In addition to these proto-words and their descendant forms, the language families have inherited word-forms for boabs. Worrorran (Northern) has the forms *potkurri* and *wajarr* in the Wunambal language; is restricted to this specific language, while the latter is also used in Wurla (Eastern Worrorran). *Wajarr* has either borrowed into the Bunuban family or from it: evidence is equivocal at this stage. Forms of the Jarragan proto-word *jumulu* have been borrowed into northern Worrorran and into the **PNy** Ngumpin subgroup. Jarragan also has the term *kertewun* that has been borrowed further east into the **nPNy** Mirndi language family.

Of particular interest is the boab word-form *larrkarti*, which is from a language subgroup associated with the desert of the main bioregions of *A. gregorii* distribution. Analyzability of this word into two morphemes (*larr*=split; *karti*=side) suggests it comes from the Karajarri (and/or the closely related and neighbouring Mangarla) languages of the **PNy** Marrngu southwest periphery of the Kimberley and is of relatively recent origin [53]. While both the elements *larr* 'split length' and *karti* 'towards; side' do occur in other languages in the region separately and with somewhat different meanings, the coincidence with Karajarri and Mangarla languages is a strong indication that this is the origin of the new coined term. As a whole, *larrkarti* means 'split side', which could be an allusion to the hollow trunks of old trees or the dehiscent boab fruit pod, or more likely to the manner in which the shelled fruit splits into longitudinal segments. The coining of the word well have come about when the Marrngu speaking people moved from the desert into the west Kimberley areas where they have encountered these unfamiliar trees in the landscape. This Marrngu word for boab has been borrowed and included into the vocabularies of some of the **nPNy** Worrorran (Western and Eastern) – Bunuban – Nyulnyulan language families. Jarragan language alongside other inherited word-forms. *Larrkarti* has also been borrowed into the **PNy** Ngumpin language subgroups from desert areas southeast of the Kimberley.

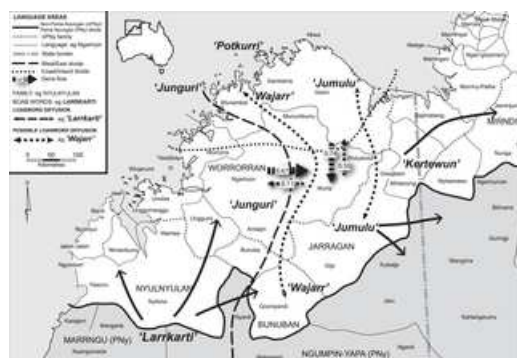
#### Boab word-forms and genetic population phylogeny

Fig. 2B illustrates the overlap between *A. gregorii* words-forms and population phylogeny. The NC population, which is sister to the remaining populations, has the inherited Worrorran boab word-forms *junguri* and *potkurri*, and possibly the NWC population has the inherited form *junguri*. The WK population, which clusters with NWC and is distributed across the **nPNy** language family areas has three words, *junguri*, *wajarr*, and *larrkarti*. The NEC population has *wajarr*, *jungule* (cognate of *junguri*) and the Jarragan loanword *jumulu*. The EK population, which occurs across the Jarragan and Nyulnyulan language families has the word-forms *jumulu* and *kertewun*. Each pair of genetic clusters shares at least one boab word-form suggesting word exchange.

#### Gene flow and diffusion of boab loanwords

Statistically significant gene flow was recorded across all boab populations in the Kimberley [40]. Migration rates were calculated by grouping the populations across two axes: West (WK, NWC) ↔ East (NC, NEC, EK); and Coast (NWC, NC, NEC) ↔ Inland (WK, EK). The West→East migration rate was 0.47 individuals/ year and the East→West rate was 0.11. The Coast→Inland rate was 0.74, and Inland→Coast was 0.50 [40].

We compared these gene flow patterns with inferred loanword diffusion patterns (Fig. 3). The linguistic analysis of loanword diffusion is consistent with the existence of an East-West biogeographic divide [36–39, 40]: *jumulu* and *kertewun* have diffused further to the east, but not into west Kimberley, whereas the word *junguri* remains in the northwest and has not diffused into east Kimberley. *Larrkarti* has diffused widely from its inferred source in the southwest into western and central Kimberley but not further into northern Worrorran languages.



**Fig 3. Gene flow between *A. gregorii* populations and boab loanword diffusion across the Kimberley region.**

Approximate locations of inferred East-West and Coast-Inland biogeographic divides are represented by dotted lines.



flow between East-West and Coast-Inland is calculated using IMa2 is in units of individuals/year, and shown with block arrows. Unbroken arrows show the direction of borrowing of boab loanwords among the Kimberley languages, and broken arrows show possible directions of loanword movements.  
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If human-mediated dispersal were responsible for the East ↔ West and Coast ↔ Inland dispersal, we might expect corresponding loanword diffusion along these axes. Furthermore, since there is evidence of much more west-to-east than east-to-west gene flow across the inferred East-West divide, we might expect more west-to-east diffusion of loanwords than the reverse. These directional patterns accord well with the diffusion of boab loanwords shown in Fig. 3. *Larrkarti* and *Ngindim* have diffused West → East, but there are no cases of East → West boab loanword diffusion. Thus, the direction and rate of migration of *A. gregorii* gene flow show correspondence with the inferred diffusion of boab loanwords.

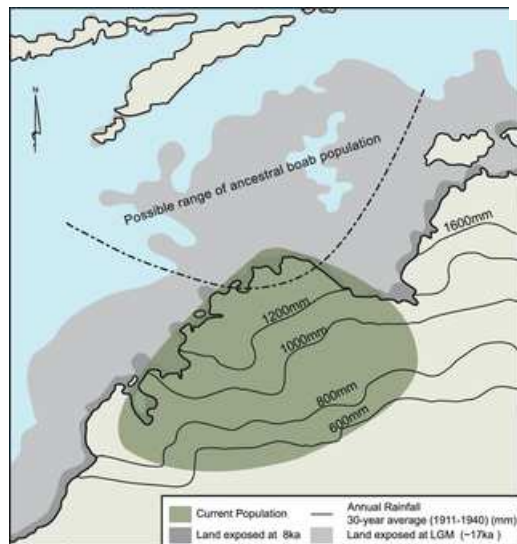
## Discussion

Our study supports the hypothesis that human-mediated dispersal has played a role in shaping the geographical distribution of *A. gregorii*. The limited morphological and genetic divergence of the *Adansonia* genus in northwest Australia, despite the presence of biogeographic barriers, can be attributed to high gene flow within *A. gregorii* [40]. Given the lack of other obvious selective agents, the lack of evidence of barriers to gene flow is most easily explained by a long history of humans moving between populations. Concordance between gene flow and loanword diffusion further supports the hypothesis of human-mediated dispersal of *A. gregorii*. Loanword movements across all boab populations provide an indication of patterns of interaction between families and *PNy* subgroups that would have influenced the geographic distribution of *A. gregorii* in the Kimberley region.

Additional evidence of anthropogenic agency in facilitating boab gene flow comes from archaeological excavations of the Gap Shelter located in the Napier ranges of the Kimberley, which falls within the Bunuba language area [48,49]. Little macrobotanical remains from this rockshelter site showed continuity of human occupation extending over 40 ka into the Pleistocene. Carbon dates recorded boab pod fragments at 39 ka, 20 ka, 18 ka and 15 ka [48,49], with substantial charcoal deposition from 3 ka onwards and peaking at around 650 years ago [49]. Old boab trees have been found near archaeological sites in western Kimberley, providing further evidence of long-term human consumption of the fruit [48]. Prehistoric rock art in northern Kimberley also shows possible depictions of the tree, indicating its cultural significance to ancient human groups that have occupied this region [28].

Based on the above evidence, we postulate that recent boab evolution and geographic distribution have been shaped through ancient human agency. The phylogenetic tree of boab populations and predominant direction of gene flow indicate that the source populations for *A. gregorii* dispersal were most likely in the extreme northwest Kimberley, potentially overlapping with the inferred NC population area. The range of this source population is likely to have extended beyond the current coastline during the Last Glacial Maximum (LGM, roughly 20 ka) when sea levels were over 120 m below present-day, and the northern continental shelf was exposed to the maximum extent [68]. The increased land surface exposure of both Sahul and the Australian continental shelves, lower sea surface temperatures (SST), and altered oceanic currents due to closure of several shallow seas and passageways between these continental shelves contributed to a northward shift of the Inter-Tropical Convergence Zone thereby reducing seasonal precipitation levels [69] and creating semi-arid savanna conditions [70] in which ancestral *A. gregorii* populations would have existed in northwest Australia.

Fig. 4 sketches a possible LGM scenario for this boab population distribution, showing land exposed at 17 ka and a comparison to present-day coastlines. With rainfall as much as 30 to 50% below present day levels [69] and higher levels of aridity [71], subtropical desert conditions would have prevailed across much of the exposed continental shelf beyond the Kimberley [72–75]. These arid climatic conditions, particularly the low levels of seasonal rainfall, would have limited the distribution of *A. gregorii*, as the current distribution of the species coincides with areas receiving at least 700 mm of annual rainfall [76]. Therefore, under drier climatic conditions during the LGM due to the northward shift of the ITCZ [69], it is likely that *A. gregorii* populations would have been limited to the extreme northern coast of present-day Kimberley and the exposed continental shelf.



**Fig 4. Sketch of LGM scenario showing land exposed at 17 ka and at 8 ka in relation to present-day coastlines, based on Collier [1].** The possible extent of ancestral boab population distribution on the exposed continental shelf is shown by a dashed line. The current boab distribution is shown in light grey. Rainfall isohyets (mm) are based on 30-year annual average rainfall (1911–1940) estimates obtained from the Australian Bureau of Meteorology [88].  
doi:10.1371/journal.pone.0119758.g004

Subsequent sea-level rise and restoration of monsoonal activity during the post-glacial period and the Pleistocene-Holocene transition between 17 and 6 ka flooded the Sahul shelf and established the present-day coastlines of northern Australia. The flooding of the continental shelf beyond the present-day Kimberley coast, along with increased monsoonal rainfall in Kimberley, would have altered the distribution of ancestral *A. gregorii* populations and possibly created a genetic bottleneck, which current populations would have expanded [40]. We propose that ancient human groups that lived on the coast previously exposed shelf during the Late Pleistocene would have retreated from inundated areas and carried boab seeds as they migrated further south and east.

Late Pleistocene records of boab remnants at the Carpenter's Gap archaeological site [48,49] may represent sporadic human settlers from the north [77]. The site record of the presence of shells and beads from the Early Holocene suggests movement of high value goods from the coast [78]. In common with other Australian archaeological sites [79], evidence of occupation at Carpenter's Gap increases sharply from mid- to late Holocene, perhaps reflecting a demographic expansion in southern Kimberley [79]. This population increase, combined with other factors such as climatic and vegetation change, led to growth in local boab populations, and more frequent use of the site for cultural ceremonies and exchanges [77] could increase in boab pod remnants along with other food plants and seeds from about 3000 to 650 years ago [48].

The increased mobility of Aboriginal groups in the southern Kimberley during the Late Holocene is likely to have been greater due to climate variability in northern Australia. In contrast to the Early Holocene, the Late Holocene (~1000 BCE – present) in the Australian Monsoon Tropics was marked by periods of increased seasonality and aridity [80–82]. These conditions contributed to increased mobility of Aboriginal groups between different parts of the Kimberley and would have contributed to *A. gregorii* gene flow through fruit and seed dispersal, accompanied by diffusion of associated word-forms.

The linguistic data provides additional indication of patterns of migration and social interaction that could have contributed to the spread of *A. gregorii* (Table 1 and Fig. 3). The periods of aridity during the Late Holocene may have affected the survival of desert-based Marrngu (**PNy** subgroup) speakers and led to their migration into southern and western Kimberley. The word *larrkarti* was probably coined by Marrngu speakers during this period when they would have encountered the boab in the Kimberley landscape. At the same time, the mobility and cultural interactions of these **PNy** groups with neighbouring language groups such as Nyikina and Bunuba would have increased. Evidence that the diffusion of the loanword *larrkarti* is of a relatively recent nature is demonstrated in the way it has been incorporated into other languages, generally in an unmodified form [61]. The Gija language (southern Jarragan) has retained its inherited word *jumulu* for the tree and adopted *larrkarti* for the fruit, perhaps indicating the salience and portability of the edible seed pod in the more recent borrowing. Likewise, the Jaru (**PNy** Ngumpin subgroup) has no inherited words for boab and uses both *jamula* (modified from Gija) and *larrkarti* for the fruit. Other examples of recent boab expansion and loanword diffusion further east can be found in the Ngarinyman language in the Northern Territory, where the words *jang-nge* (borrowed from Miriwoong, meaning 'for eating') and *jumulu* (from Gija) are used for the fruit or its edible pith, and the tree respectively.

The possible climatic influences on human migration and boab loanword movement in the Kimberley echoes some of Bostoen et al.'s [6] description of climate-induced dynamics and Bantu expansion in Africa. Although there is no archaeological evidence of a large-scale expansion of a single linguistic group into the Kimberley, the **PNy** Marrngu term *larrkarti* and pre-existing **nPNy** language words such as *wajarr*, *jumulu*, and *kertewun* moved across the Kimberley in patterns consistent with



with multi-directional baob gene flow as shown in our study.

The high gene flow in *A. gregorii* appears similar to the case of *A. digitata* in Africa, where human agency has been dispersal of the species [19,51,52]. The baob loanword movements in the Kimberley may be compared with Blench the spread of Bantu words for *A. digitata*. He notes that the genetic diversity of baobabs in the ecological zones of the diversity of vernacular names for the tree in African languages suggests considerable antiquity as well as significant movement along trade routes and exchanges of associated ideas and terminology. However, despite the diversity of names, he points out that two competing Bantu roots, *#mbuyu* and *#muramba*, and variations of these, are found in languages of southern and eastern Africa. The Bantu expansion from the tropical forest areas of West Africa is said to have occurred around the middle of the first millennium BC (~ 2500 BP onwards) and reached southern Africa by about 500 AD. Blench argues that the Bantu would not have been familiar with the baobab because it does not grow in the tropical forest of Cameroon, Gabon and Congo where this proto-language group are thought to have originated. They would have encountered the tree as they expanded eastwards and emerged into the savanna, and developed new terms by either borrowing from hunter-gatherer groups or comparing it with some other tree species they already knew. The loan words or variations of *mbuyu* and *muramba* in the Bantu languages of eastern and southern Africa would thus indicate the movement of Bantu into these areas [4]. The lower levels of genetic diversity of *A. digitata* in eastern and southern Africa detected by Leong Pock may likely be due to the Bantu expansion over this 3000 year period and their contribution to high levels of gene flow across these regions.

#### Conclusion

Our study demonstrates that the limited intraspecific divergence within *A. gregorii* in Australia is most likely due to human agency, similar to that inferred for *A. digitata* in continental Africa [27, 51], combined with shifts in habitat and a weak bottleneck following the end of the LGM [40]. Human use of *Adansonia* over many thousands of years on both continents would have contributed to gene flow over long distances and across biogeographical barriers. In contrast, the divergence of *Adansonia* into six species in Madagascar was possible in part because of the lack of humans until recently. However, this hypothesis can only be tested by further investigation of the ecological, physiological and biogeographical factors contributing to speciation within the *Adansonia* clade from Madagascar.

This study contributes new evidence for the role of ancient humans in influencing the evolution and distribution of native plant species in Australia. Australia has long been viewed as a continent of hunter-gatherers [84] where prehistoric populations prior to European colonisation played minimal roles in selecting and dispersing useful plants [23,24]. However, recent studies have challenged this assumption by providing evidence for the role of ancient Aboriginal groups in the food plants across the continent [77,85,86]. These include bananas (*Musa* spp.), taro (*Colocasia esculenta*) and soya (*Dioscorea* spp.) in northern Australia [23], *Livistona* palms in Central Australia [85], and yam daisy (*Microseris scabra*) in southeastern Australia [86]. Our findings add new insights regarding the role of ancient human agency in influencing the distribution of the baob, an important non-cultivated food plant species that has shaped the long-term landscape and environmental history of northwest Australia.

#### Supporting Information

**S1 Dataset. Genetic data from six nuclear microsatellite loci of 220 *A. gregorii* individuals.**

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(XLS)

**S2 Dataset. Terms for baob in Aboriginal languages of the Kimberley region.**

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#### Author Contributions

Conceived and designed the experiments: KLB RF DJM. Performed the experiments: KLB RF. Analyzed the data: KLB RF TS SS. Contributed reagents/materials/analysis tools: KLB HR. Wrote the paper: HR KLB DAB. Conceived the overall project: HR DJM CAK. Analysed the genetic data: KLB RF DAB. Contributed collection of baob terms in Aboriginal languages of the Kimberley region and analysed the linguistic data: PM TS SS.

#### References

1. Donohue M, Denham T. Banana (*Musa* spp.) domestication in the Asia-Pacific region: linguistic and archaeobotanical perspectives. E

Research and Applications. 2009;7: 293.

[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)

2. Perrier X, De Langhe E, Donohue M, Lentfer C, Vrydaghs L, Bakry F, et al. Multidisciplinary perspectives on banana (*Musa* spp.) dom  
Acad Sci USA. 2011;108(28): 11311–8. doi: 10.1073/pnas.1102001108. pmid:21730145  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
3. Roullier C, Benoit L, McKey DB, Lebot V. Historical collections reveal patterns of diffusion of sweet potato in Oceania obscured by mo  
movements and recombination. Proc Natl Acad Sci USA. 2013;110: 2205–10. doi: 10.1073/pnas.1211049110. pmid:23341603  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
4. Blench RM. The intertwined history of the silk-cotton and baobab. In: Cappers RTJ, editor. Fields of change; progress in African ethno  
Germany: Barkhuis and Gröningen University, Gröningen; 2007. p. 1–20.
5. Smith BD. General patterns of niche reconstruction and the management of 'wild' plant and animal resources by small-scale pre-indus  
Philosophical Transactions of the Royal Society B: Biological Sciences. 2011;366: 838–48. doi: 10.1098/rstb.2010.0253. pmid:217271  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
6. Bostoen K, Grollemund R, Muluwa JK. Climate-induced vegetation dynamics and the Bantu Expansion: Evidence from Bantu names 1  
(*Elaeis guineensis*, *Canarium schweinfurthii*, and *Musanga cecropioides*). Comptes Rendus Geoscience. 2013;345: 336–49. doi:  
10.1016/j.crte.2013.03.005  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
7. Baum DA. A systematic revision of *Adansonia* (Bombacaceae). Ann Mo Bot Gard. 1995;82: 440–71. doi: 10.2307/2399893  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
8. Pettigrew DJ, Bell KL, Bhagwandin A, Grinan E, Jillani N, Meyer J, et al. Morphology, ploidy and molecular phylogenetics reveal a nev  
from Africa in the baobab genus *Adansonia* (Bombacoideae; Malvaceae). Taxon. 2012;61: 1240–50.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
9. Adanson M. Description d'un arbre d'un nouveau genre, appelé baobab, observé au Sénégal [Description of a new tree of a new gen  
observed in Senegal]. Mémoires de l'Académie Royale. 1771;161: 218–43.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
10. De Caluwé E, Halamová K, Van Damme P. Boabab (*Adansonia digitata* L.): a review of traditional uses, phytochemistry and pharmaci  
Simon JE, Ho C-T, editors. African natural plant products: new discoveries and challenges in chemistry and quality. Washington DC: A  
Society; 2009. p. 51–84.
11. Gebauer J, El-Siddig K, Ebert G. Baobab (*Adansonia digitata* L.): a review on a multipurpose tree with promising future in the Sudan.  
Gartenbauwissenschaft. 2002;67: 155–60.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
12. Livingstone D. A popular account of missionary travels and researches in South Africa. London: John Murray, Albemarle Street; 1861.
13. Patrut A, von Reden KF, Mayne DH, Lowy DA, Patrut RT. AMS radiocarbon investigation of the African baobab: Searching for the old  
Instrum Methods Phys Res B. 2013;294: 622–6. doi: 10.1016/j.nimb.2012.04.025.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
14. Sidibe M, Williams JT. Baobab. *Adansonia digitata*. Southampton, UK: International Centre for Underutilised Crops; 2002.
15. Swart ER. Age of the baobab tree. Nature. 1963;4881: 708–9. doi: 10.1038/198708b0  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
16. Wickens GE. The baobab: Africa's upside-down tree. Kew Bulletin. 1982;37: 173–209. doi: 10.2307/4109961  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
17. Wickens GE, Lowe P. The baobabs: pachycauls of Africa, Madagascar, and Australia. Berlin; New York: Springer; 2008.
18. Armstrong P, editor. The history, natural history and distribution of *Adansonia*: a plant genus of the Indian Ocean littoral. The Indian Oc  
International Conference on Indian Ocean Studies, Perth, Western Australia 1979: Section I Environment and Resources; 1979; Perth  
Perth Building Society.

19. Duvall CS. Human settlement and baobab distribution in south-western Mali. *Journal of Biogeography*. 2007;34: 1947–61. doi: 10.1111/j.1365-2699.2007.01751.x.  
View Article • PubMed/NCBI • Google Scholar
20. Barker G. *The Agricultural Revolution in Prehistory*. Oxford: Oxford University Press; 2006.
21. Bean AR. A new system for determining which plant species are indigenous in Australia. *Australian Systematic Botany*. 2007;20: 1–43. doi: 10.1071/SB06030.  
View Article • PubMed/NCBI • Google Scholar
22. Bellwood P. *First Farmers*. Oxford: Blackwell; 2005.
23. Denham T, Donohue M, Booth S. Revisiting an old hypothesis: Horticultural experimentation in northern Australia. *Antiquity*. 2009;83: 10.1017/S0003598X00098884  
View Article • PubMed/NCBI • Google Scholar
24. Lourandos H. *Continent of hunter-gatherers: New perspectives in Australian prehistory*. Cambridge: CUP; 1997.
25. Bliege-Bird R, Bird DW, Coddling BF, Jones JH. The “fire stick farming” hypothesis: Australian Aboriginal foraging strategies, biodiversity and anthropogenic fire mosaics. *Proc Natl Acad Sci USA*. 2008;105: 14796–801. doi: 10.1073/pnas.0804757105. pmid:18809925  
View Article • PubMed/NCBI • Google Scholar
26. Jones R. Firestick farming. *Australian Natural History*. 1969;16: 224–31.  
View Article • PubMed/NCBI • Google Scholar
27. Leong Pock Tsy J-M, Lumaret R, Mayne D, Vall AOM, Abutaba YIM, Sagna M, et al. Chloroplast DNA phylogeography suggests a West African origin for the baobab, *Adansonia digitata* L. (Bombacoideae, Malvaceae). *Molecular Ecology*. 2009;18(8): 1707–15. doi: 10.1111/j.1365-294X.2009.04144.x. pmid:19302348  
View Article • PubMed/NCBI • Google Scholar
28. Pettigrew DJ. Iconography in Bradshaw rock art: breaking the circularity. *Clin Exp Optom*. 2011;94: 403–17. doi: 10.1111/j.1444-0938.2011.02188.4255  
View Article • PubMed/NCBI • Google Scholar
29. Baum DA, Handasyde T. The baobab tree (*Adansonia gregorii*) in north-west Australia. Perth: Unpublished report in Western Australian Museum; 1990.
30. Brock J. *Top End Native Plants*. Darwin: Brock; 1988.
31. Gillison AN. Tropical savannas of Australia and southwest Pacific. In: Bourliere F, editor. *Tropical savannas*. Amsterdam: Elsevier Science; 1983. p. 183–243.
32. Mueller Fv. Botanical notes from north-west Australia. *Victorian Naturalist*. 1893;10: 110–1.  
View Article • PubMed/NCBI • Google Scholar
33. Bowman DMJS. Observations on the demography of the Australian baobab (*Adansonia gregorii*) in the north-west of the Northern Territory. *Australian Journal of Botany*. 1997;45: 893–904. doi: 10.1071/bt96092  
View Article • PubMed/NCBI • Google Scholar
34. Bowman DMJS, Brown GK, Braby MF, Brown JR, Cook LG, Crisp MD, et al. Biogeography of the Australian monsoon tropics. *Journal of Biogeography*. 2010;37(2): 201–16. doi: 10.1111/j.1365-2699.2009.02210.x.  
View Article • PubMed/NCBI • Google Scholar
35. Byrne M, Yeates DK, Joseph L, Kearney M, Bowler J, Williams MA, et al. Birth of a biome: insights into the assembly and maintenance of an arid zone biota. *Mol Ecol*. 2008;17(20): 4398–417. doi: 10.1111/j.1365-294X.2008.03899.x. pmid:18761619  
View Article • PubMed/NCBI • Google Scholar
36. Eldridge MDB, Potter S, Cooper SJB. Biogeographic barriers in north-western Australia: an overview and standardisation of nomenclature. *Journal of Biogeography*. 2011;38: 270–2. doi: 10.1071/ZO12012.  
View Article • PubMed/NCBI • Google Scholar

37. Hill KD, Johnson LAS. Systematic studies in the eucalypts. 7. A revision of the bloodwoods, genus *Corymbia* (Myrtaceae). *Telopea*. 1991;19: 1–10. doi: 10.1071/tp91001. View Article • PubMed/NCBI • Google Scholar
38. Oliver PM, Adams M, Doughty P. Molecular evidence for ten species and Oligo-Miocene vicariance within a nominal Australian gecko (*Crenadactylus ocellatus*, Diplodactylidae). *Bmc Evolutionary Biology*. 2010;10: 386. doi: 10.1186/1471-2148-10-386. pmid:20812123. View Article • PubMed/NCBI • Google Scholar
39. Potter S, Eldridge MDB, Taggart DA, Cooper SJB. Multiple biogeographical barriers identified across the monsoon tropics of northern Australia: a phylogeographic analysis of the *brachyotis* group of rock-wallabies. *Mol Ecol*. 2012;21: 2254–69. doi: 10.1111/j.1365-294X.2012.05522.x. View Article • PubMed/NCBI • Google Scholar
40. Bell KL, Rangan H, Fowler R, Kull CA, Pettigrew JD, Vickers CE, et al. Genetic diversity and biogeography of the boab *Adansonia digitata* (Bombacoideae). *Australian Journal of Botany*. 2014; 62: 164–74. doi: 10.1071/bt13209. View Article • PubMed/NCBI • Google Scholar
41. Baum DA. The comparative pollination and floral biology of baobabs (*Adansonia*—Bombacaceae). *Ann Mo Bot Gard*. 1995;82: 322–44. doi: 10.2307/2399883. View Article • PubMed/NCBI • Google Scholar
42. Lowe P. The boab tree. Port Melbourne: Lothian Books; 1998.
43. Aldrich PR, Hamrick JL, Chavarriaga P, Kochert G. Microsatellite analysis of demographic genetic structure in fragmented populations of *Symphonia globulifera*. *Mol Ecol*. 1998;7(8): 933–44. pmid:9711860 doi: 10.1046/j.1365-294x.1998.00396.x. View Article • PubMed/NCBI • Google Scholar
44. Garcia C, Jordano P, Godoy JA. Contemporary pollen and seed dispersal in a *Prunus mahaleb* population: patterns in distance and direction. *Ecology*. 2007;16(9): 1947–55. pmid:17444903 doi: 10.1111/j.1365-294x.2006.03126.x. View Article • PubMed/NCBI • Google Scholar
45. Oddou-Muratorio S, Petit RJ, Le Guerouet B, Guesnet D, Demesure B. Pollen versus seed-mediated gene flow in a scattered forest tree species. *Evolution*. 2001;55: 1123–35. pmid:11475048 doi: 10.1111/j.0014-3820.2001.tb00632.x. View Article • PubMed/NCBI • Google Scholar
46. Potter S, Eldridge MDB, Cooper SJB, Paplinska JZ, Taggart DA. Habitat connectivity, more than species' biology, influences genetic diversity of a habitat specialist, the short-eared rock-wallaby (*Petrogale brachyotis*). *Conserv Genet*. 2012;13: 937–52. doi: 10.1007/s10592-012-0312-2. View Article • PubMed/NCBI • Google Scholar
47. Boland DJ, Brooker MIH, Chippendale GM, Hall N, Hyland BPM, Johnston RD, et al. Forest trees of Australia. Melbourne: Nelson & Phillips; 2004.
48. McConnell K, O'Connor S. 40,000 year record of food plants in the Southern Kimberley Ranges, Western Australia. *Australian Archaeology*. 2003;67: 20–31. View Article • PubMed/NCBI • Google Scholar
49. McConnell K, O'Connor S. Carpenter's Gap Shelter I: A case for total recovery. In: Mountain MJ, Bowdery D, editors. *Taphonomy: the processes from phytoliths to megafauna*. Canberra, Australia: ANH Publications; 1999. p. 23–34.
50. Wallis LA. Environmental history of northwest Australia based on phytolith analysis at Carpenter's Gap 1. *Quat Int*. 2001;83–85: 103–11. doi: 10.1016/s1040-6182(01)00033-7. View Article • PubMed/NCBI • Google Scholar
51. Assogbadjo AE, Glele Kakai R, Kyndt T, Sinsin B. Conservation genetics of baobab (*Adansonia digitata* L.) in the parklands agroforests (West Africa). *Not Bot Horti Agrobot Cluj Napoca*. 2010;38: 136–40. View Article • PubMed/NCBI • Google Scholar
52. Leong Pock Tsy JM, Lumaret R, Mayne D, Vall AO, Abutaba YI, Sagna M, et al. Chloroplast DNA phylogeography suggests a West African origin for the baobab, *Adansonia digitata* L. (Bombacoideae, Malvaceae). *Mol Ecol*. 2009;18: 1707–15. doi: 10.1111/j.1365-294X.2009.03902.x. pmid:19302348. View Article • PubMed/NCBI • Google Scholar
53. McConvell P, Spronck S, Saunders T, editors. Linguistic prehistory of the Australian boab. The 43rd meeting of the Australian Linguistic Society; 2009.

Conference Proceedings; 2014.

54. Cavalli-Sforza LL, Edwards AWF. Phylogenetic analysis: models and estimation procedures. *Am J Hum Genet.* 1967;19: 233–57. pmid:10.2307/2406616  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
55. Felsenstein J. PHYLIP: Phylogeny Inference Package. 3.69 ed. Seattle, WA, USA: Department of Genome Sciences and Department University of Washington; 2009.
56. Takezaki N, Nei M. Genetic distances and reconstruction of phylogenetic trees from microsatellite DNA. *Genetics.* 1996;144: 389–99.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
57. Bowern C, Koch H. *Australian Languages: Classification and the Comparative Method.* Amsterdam: John Benjamins Publishing Company; 2003.
58. Harvey MD, cartographer Non-Pama-Nyungan Languages: Mapping Database and Maps: The Aboriginal Studies Electronic Data Archive; 2008.
59. Evans N. The Non-Pama–Nyungan Languages of Northern Australia. *Comparative studies of the continent's most linguistically complex languages.* Pacific Linguistics; 2003.
60. Campbell L. *Historical Linguistics.* Cambridge, MA: MIT Press; 1999.
61. McConvell P, Laughren M. Ngumpin-Yapa Languages. In: Koch H, Bowern C, editors. *Australian Languages: Classification and the Comparative Method.* Amsterdam: John Benjamins Publishing Company; 2004.
62. Hudson DH, Bryant D. Application of phylogenetic networks in evolutionary studies. *Mol Biol Evol.* 2006;23: 254–67. pmid:16221896  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
63. McGregor W, Rumsey A. Worrorran revisited: the case for genetic relations among languages of the Northern Kimberley region of Western Australia. *Pacific Linguistics.* 2009;L600: 14.
64. McGregor W. *The Languages of the Kimberley, Western Australia.* Abingdon, UK: Routledge Curzon; 2004.
65. Excoffier L, Smouse PE, Quattro JM. Analysis of molecular variance inferred from metric distances among DNA haplotypes: Application to mitochondrial DNA restriction data. *Genetics.* 1992;131: 479–91. pmid:1644282  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
66. Meirmans PG. The trouble with isolation by distance. *Mol Ecol.* 2012;21: 2839–46. doi: 10.1111/j.1365-294X.2012.05578.x. pmid:22571000  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
67. Maddison WP, Maddison DR. Mesquite: a modular system for evolutionary analysis. version 2.75 2009.
68. Kershaw P. Environmental change in Greater Australia. *Antiquity.* 1995;69.265: 656–75.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
69. De Deckker P, Tapper NJ, van der Kaars S. The status of the Indo-Pacific Warm Pool and adjacent land at the Last Glacial Maximum. *Quaternary Science Reviews.* 2002;21: 25–35. doi: 10.1016/S0921-8181(02)00089-9  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
70. Bird MI, Taylor D, Hunt C. Palaeoenvironments of insular Southeast Asia during the Last Glacial Period: A savanna corridor in Sundaland. *Quaternary Science Reviews.* 2005;24: 2228–42. doi: 10.1016/j.quascirev.2005.04.004  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
71. Bowler JM, Wyrwoll K-H, Lou Y. Variations of the northwest Australian summer monsoon over the last 300,000 years: the paleohydrological Gregory (Mulan) Lakes System. *Quaternary International.* 2001;83–85: 63–80. doi: 10.1016/S1040-6182(01)00031-3  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
72. Reeves JM, Bostock HC, Ayliffe LK, Barrows TT, De Deckker P, Devriendt LS, et al. Palaeoenvironmental change in tropical Australia 30,000 years—a synthesis by the OZ-INTIMATE group. *Quaternary Science Reviews.* 2013;74: 97–114. doi: 10.1016/j.quascirev.2012.11.011  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
73. Hesse PP, Magee JW, van der Kaars S. Late Quaternary climates of the Australian arid zone: a review. *Quaternary International.* 2004;124: 1–14.

doi: 10.1016/s1040-6182(03)00132-0

[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)

74. Fitzsimmons KE, Cohen TJ, Hesse PP, Jansen J, Nanson GC, May J-H, et al. Late Quaternary palaeoenvironmental change in the Australian Quaternary Science Reviews. 2013;74: 78–96. doi: 10.1016/j.quascirev.2012.09.007.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
75. Wyrwoll K-H, Miller GH. Initiation of the Australian summer monsoon 14,000 years ago. Quaternary International. 2001;83–85: 87–100. doi: 10.1016/s1040-6182(01)00034-9  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
76. Beard JS. Some vegetation types of tropical Australia in relation to those of Africa and America. Journal of Ecology. 1967;55(2): 271–287. doi: 10.2307/2257877  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
77. McConnell K. Palaeoethnobotanical remains of Carpenter's Gap Site 1, The Kimberleys, Western Australia. MSc Thesis. Canberra: Australian Archaeology. 2014;78: 10–23.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
78. O'Connor S, Maloney T, Vannieuwenhuysen D, Balme J, Wood R. Occupation at Carpenters Gap 3, Windjana Gorge, Kimberley, Western Australia. Australian Archaeology. 2014;78: 10–23.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
79. Williams AN. A new population curve for prehistoric Australia. Proceedings of the Royal Society B: Biological Sciences. 2013;280: 20130486. doi: 10.1098/rspb.2013.0486. pmid:23615287  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
80. Lees BG, Clements A. Climatic implications of chenier dates in northern Australia. Radiocarbon. 1987;29: 311–7.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
81. Brockwell C, Marwick B, Bourke P, Faulkner P, Willan R. Late Holocene climate change and human behavioural variability in the coastal northern Australia: Evidence from a pilot study of oxygen isotopes in marine bivalve shells from archaeological sites. Australian Archaeology. 2013;77: 21–33.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
82. Shulmeister J. Australasian evidence for mid-Holocene climate change implies precessional control of Walker Circulation in the Pacific. Quaternary International. 1999;57: 81–91. doi: 10.1016/s1040-6182(98)00052-4  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
83. Ehret C. Bantu Expansions: Re-envisioning a central problem of early African history. The International Journal of African Historical Studies. 2009;41: 5–41. doi: 10.2307/3097285  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
84. Denham T, Fullagar R, Head L. Plant exploitation on Sahul: From colonisation to the emergence of regional specialisation during the Holocene. Quaternary International. 2009;202: 29–40. doi: 10.1016/j.quaint.2008.06.018  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
85. Kondo T, Crisp MD, Linde C, Bowman DM, Kawamura K, Kaneko S, et al. Not an ancient relic: the endemic *Livistona* palms of arid Australia have been introduced by humans. Proc Biol Sci. 2012;279(1738): 2652–61. Epub 2012/03/09. doi: 10.1098/rspb.2012.0103 pmid:22312223 Central PMCID: PMC3350701.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
86. Gott B. Murnong—*Microseris scapigera*: a study of a staple food of Victorian Aborigines. Australian Aboriginal Studies. 1983;2: 2–17.  
[View Article](#) • [PubMed/NCBI](#) • [Google Scholar](#)
87. Collier M. Sahul Time. Available from: <http://sahultime.monash.edu.au/explore.html>, accessed 5 April 2014
88. Australian Bureau of Meteorology. Available: [http://www.bom.gov.au/jsp/ncc/climate\\_averages/decadal-rainfall/index.jsp?maptype=6&product=totals-maps](http://www.bom.gov.au/jsp/ncc/climate_averages/decadal-rainfall/index.jsp?maptype=6&product=totals-maps). Accessed 2014 April 5.